Discovery of DS-1558: A Potent and Orally Bioavailable GPR40 Agonist

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Supporting Information

ABSTRACT: GPR40 is a G protein-coupled receptor that is predominantly expressed in pancreatic β -cells. GPR40 agonists stimulate insulin secretion in the presence of high glucose concentration. On the basis of this mechanism, GPR40 agonists are possible novel insulin secretagogues with reduced or no risk of hypoglycemia. The improvement of *in vitro* activity and metabolic stability of compound 1 led to the discovery of 13, (3S)-3-ethoxy-3-(4-{[(1R)-4-(trifluoromethyl)-2,3-dihydro-1H-inden-1-yl]oxy}phenyl)propanoic acid, as a potent and orally available GPR40 agonist. Compound 13 (DS-1558) was found to have potent glucose lowering effects during an oral glucose tolerance test in ZDF rats.



KEYWORDS: GPR40, agonist, insulin secretagogue, diabetes, glucose lowering

T ype 2 diabetes is a metabolic disorder characterized by impaired glucose homeostasis caused by insufficient insulin secretion or insulin resistance. The number of diabetics has been increasing all over the world and has reached nearly 350 million.^{1,2} Current common therapies include the use of insulin injections, sulfonylureas, metformin, and glinides.^{3,4} Most of them are associated with problems such as weight gain, risk of hypoglycemia, and lack of sustained efficacy.^{5–9} Lately, glucose-dependent insulin secretagogues, such as GPR119¹⁰ and GPR142¹¹ agonists, have attracted attention as alternative treatments for diabetes. Among them, DPP-IV inhibitors enhancing the activity of GLP-1 have already been widely used. Most recently, SGLT2 inhibitors that inhibit the reuptake of urinal sugar have been developed as hypoglycemic agents with low risk of weight gain.

GPR40 is primarily expressed in pancreatic β -cells and activated by long-chain free fatty acids, resulting in enhancement of glucose-stimulated insulin secretion (GSIS) dependent on elevated glucose levels.^{12–17} On the basis of this GSIS mechanism, GPR40 has also received considerable attention as a novel therapeutic target for type 2 diabetes because of its low risk of hypoglycemia.^{18,19} Recently, several groups have reported GPR40 agonists that contain acidic moieties such as a carboxylic acid or thiazolidinedione (Figure 1).^{20–28} We have also identified 3-ethoxypropanoic acid 1 as a promising lead compound (Figure 2).²⁹

Herein we describe the lead optimization of **1** to discover (3S)-3-ethoxy-3- $(4-\{[(1R)-4-(trifluoromethyl)-2,3-dihydro-1H-inden-1-yl]oxy\}$ phenyl)propanoic acid (13), a potent and orally available GPR40 agonist.



Figure 1. Reported GPR40 agonists.^{20–28}



Figure 2. Optimization of lead compound 1.

Although compound 1 showed a glucose lowering effect in rats after oral administration, its half-life was very short (Table 2). High *in vivo* clearance of 1 was probably due to metabolic oxidation at the benzyl position because we detected a glutathione (GSH) adduct of 2-methylbenzaldehyde, a putative metabolite of 1, from a GSH trapping assay in human liver microsomes. Therefore, we designed the cyclized compounds

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between the benzylic position and the ortho position of the phenyl ring to avoid benzyl oxidation (Figure 2).

First we investigated the stereoconfiguration of the ethoxy moiety (Table 1). Chiral separation of the Cl-substituted derivative revealed that (S)-stereochemistry was preferred for GPR40 agonistic activity (2-4).

Next we synthesized cyclized derivatives with 5- or 6membered rings. Among indane derivatives (6-8), compound 6 was the most potent agonist. Tetralin derivative 9 with the

Table 1. Structures and in Vitro Activities of thePhenylpropanoic Acid Series

Compd	R	Configuration of β -ethoxy moiety	GPR40 hEC50 (nM)ª
1	Me	racemic	20
2		racemic	6.2
3	CI	ر (S)-	3.9
4	CI	ر ب (<i>R</i>)-	360
5	F ₃ C	(S)-	5.3
6	S.	(S)-	31
7		(S)-	69
8		(<i>S</i>)-	83
9		(<i>S</i>)-	62
10	Me	<i>(S)</i> -	29
11	Eto	(<i>S</i>)-	63
12	CI KS	<i>(S)</i> -	5.0
13 (DS- 1558)	F ₃ C	(S)-	3.7
14	F ₃ C	(S)-	19
15	F ₃ C	(R)-	330
16	F ₃ C	(R)-	2600

^{*a*}A calcium flux assay in transfected-GPR40 CHO cells. Assay results are the average of triplicates. Standard deviation was $\pm 20\%$.

same stereochemistry at the benzyl position was weaker than 6. Accordingly, we focused our investigation to the substituents on the indane ring. The methyl (10) and ethoxy (11)derivatives showed similar potency to compound 6. The Cl (12) and CF_3 (13) substituents provided a significant leap in agonistic activity. We synthesized possible diastereomers (14-16) of 13 and confirmed that compound 13 was the most potent agonist. The stereoconfiguration at both indane and ethoxy moieties significantly impacted the GPR40 agonistic activity. Furthermore, compound 13 had no PPARy agonistic activity up to 100 μ M, whereas noncyclic compound 5 had partial PPAR γ agonistic activity (EC₅₀ = 10.4 μ M, E_{max} =11.3% relative to rosiglitazone). This indicated that the fixed conformation improved the GPR40 selectivity over PPARy. In addition, 13 had no activity for GPR120 and exhibited >100fold selectivity over 68 other diverse receptors, ion channels, and transporters.³⁰ Thus, potent and selective GPR40 agonist 13, named DS-1558, was selected for further investigations.

Compound 13 (DS-1558) was synthesized as shown in Scheme 1. This route was considered to be able to synthesize



"Reagents and conditions: (i) HCO_2H , NEt_3 , RuCl[(R,R)-Tsdpen]-(mesitylene), r.t., 85%, 99% ee; (ii) 4-fluorobenzonitrile, NaH, THF/ DMF, r.t., quant.; (iii) 5 N NaOH aq., 2-methoxyethanol, 120 °C, 84%; (iv) methyl potassium malonate, CDI, MgCl₂, NEt₃, THF/ EtOAc, r.t. to 45 °C, 84%; (v) HCO_2H , NEt₃, RuCl[(S,S)-Tsdpen](mesitylene), 38 °C, 91%, 96% de; (vi) EtI, Ag₂O, toluene, 110 °C, 84%; (vii) LiAlH₄, THF, 0 °C, quant.; (viii) TEMPO, NaClO₂, NaClO, pH 6.86 phosphate buffer, acetonitrile, 0 to 10 °C, 85%, 98% de.

this compound on a large scale using mild reactions, which were arranged to suppress the production of diastereomers minimally. In actual fact, we obtained 80 g of 13 (DS-1558) starting from about 90 g of 17 with high optical purity (98% de).

Starting from a commercially available indanone 17, chiral alcohol 18 was obtained by Ru-catalyzed asymmetric transfer hydrogenation with good enantioselectivity (99% ee).³¹ A S_NAr reaction was used to obtain aryl ether 19. After yielding benzoic acid 20 on hydrolysis in an alkali solution, β -ketoester 21 was synthesized by the condensation of magnesium methylmalonate with acid imidazolide preformed from 20 and CDI.³² The chiral Ru-catalyst with opposite chirality was utilized again for

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the asymmetric hydrogenation of β -ketoester to give β hydroxyester **22** in a good diastereo ratio (96% de). The β hydroxy group in **22** was alkylated in the neutral conditions of EtI and Ag₂O without the retro-aldol reaction. The ester group of **23** was reduced into primary alcohol and subsequently oxidized to carboxylic acid **13** (DS-1558) by TEMPO. This multistep procedure was adopted in order to avoid β elimination.

The absolute stereochemistry of **13** (DS-1558) was determined by X-ray crystal structural analysis in the form with (S)-arginine and methanol solvate. On the basis of absolute configuration of the (S)-arginine, the absolute configurations of the ethoxy and the indane moieties were determined as (S) and (R), respectively (Figure 3).



Figure 3. ORTEP representation of 13 (DS-1558).

Pharmacokinetic (PK) parameters were summarized in Table 2. Compound 13 (DS-1558) showed higher plasma exposure

Table 2. Comparison of Pharmacokinetic Properties of 1 and 13 (DS-1558) in Rats, Dogs, and Monkeys^a

compd	1	13 (DS-1558)		
species	rat	rat	dog	monkey
i.v.				
dose (mg/kg)	2	1	0.5	1
Cl (L/h/kg)	0.37	0.020	0.042	0.032
$V_{\rm dss}~({\rm L/kg})$	0.16	0.19	0.23	0.62
$T_{1/2}$ (h)	0.37	6.0	4.0	17
p.o.				
dose (mg/kg)	3	1	1	1
$C_{\rm max} \ (\mu g/mL)$	3.3	3.1	1.7	2.3
AUC ($\mu g \cdot h/mL$)	5.3	40	14	30
F (%)	64	80	66	100
^{<i>a</i>} The data is the mea Supporting Information	in value.	The statistics	s are show	vn in the

and longer half-life than compound 1 in rats. The improved PK profiles were also observed in monkeys and dogs. As we expected, the cyclization approach was efficient at improving the PK profiles.

The GPR40-mediated effects of **13** (DS-1558) on glucosestimulated insulin secretion were confirmed in isolated islets from GPR40 KO and wild-type mice. Improvement of glucose tolerance and insulin secretion were also recognized during the *in vivo* studies of these mice and ZF rats.³³ Furthermore, we evaluated *in vivo* efficacy of this compound by an oral glucose tolerance test (OGTT) in Zucker diabetic fatty (ZDF) rats, which exhibit obesity with diabetes and are widely used for therapeutic research on type 2 diabetes. The compound was orally administrated 30 min prior to a glucose challenge (2 g/ kg). Even at the minimum dosing, 0.03 mg/kg, the **13** (DS-1558) treatment markedly reduced the glucose excursion compared to the control (Figure 4A). The glucose-lowering



Figure 4. Effects of **13** (DS-1558) during an OGTT in male ZDF rats. Time-dependent changes of plasma glucose (A) and plasma insulin (B). The results are represented as the mean \pm standard error (n = 6, 9-weeks old).

potency of 13 (DS-1558) at 0.1 mg/kg was similar to that of sitagliptin at 10 mg/kg. Simultaneously, the augmentation of plasma insulin levels by 13 (DS-1558) at 0.1 mg/kg and sitagliptin at 10 mg/kg was observed (Figure 4B).

In conclusion, we have shown the lead optimization of 3ethoxypropanoic acid 1 to identify the promising compound 13 (DS-1558). The cyclization of benzyl carbon improved not only the half-life but also GPR40 selectivity. We developed the chiral synthesis of 13 (DS-1558) on a large scale with Rucatalyzed asymmetric transfer hydrogenation as a key reaction. The potent *in vivo* glucose lowering effect of 13 (DS-1558) was demonstrated in ZDF rats, type 2 diabetic model rats. Further details of 13 (DS-1558) will be reported in due course.

ASSOCIATED CONTENT

Supporting Information

Experimental procedures and analytical data. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

ABBREVIATIONS

CDI, 1,1'-carbonyldiimidazole; TEMPO, 2,2,6,6-tetramethylpiperidine *N*-oxyl; PPAR, peroxisome proliferator-activated receptor

REFERENCES

(1) Danaei, G.; Finucane, M. M.; Lu, Y.; Singh, G. M.; Cowan, M. J.; Paciorek, C. J.; Lin, J. K.; Farzadfar, F.; Khang, Y. H.; Stevens, G. A.; Rao, M.; Ali, M. K.; Riley, L. M.; Robinson, C. A.; Ezzati, M. National, regional, and global trends in fasting plasma glucose and diabetes prevalence since 1980: systematic analysis of health examination surveys and epidemiological studies with 370 country-years and 2.7 million participants. *Lancet* 2011, 378, 31–40.

(2) WHO. Diabetes Fact Sheet, No. 312, October 2013.

(3) Doyle, M. E.; Egan, J. M. Pharmacological agents that directly modulate insulin secretion. *Pharmacol. Rev.* **2003**, *55*, 105–131.

(4) Seino, Y.; Rasmussen, M. F.; Nishida, T.; Kaku, K. Efficacy and safety of the once-daily human GLP-1 analogue, liraglutide, vs glibenclamide monotherapy in Japanese patients with type 2 diabetes. *Curr. Med. Res. Opin.* **2010**, *26*, 1013–1022.

(5) Maedler, K.; Carr, R. D.; Bosco, D.; Zuellig, R. A.; Berney, T.; Donath, M. Y. Sulfonylurea induced β -cell apoptosis in cultured human islets. *J. Clin. Endocrinol. Metab.* **2005**, *90*, 501–506.

(6) Del Guerra, S.; Marselli, L.; Lupi, R.; Boggi, U.; Mosca, F.; Benzi, L.; Del Prato, S.; Marchetti, P. Effects of prolonged in vitro exposure to sulphonylureas on the function and survival of human islets. *J. Diabetes Complications* **2005**, *19*, 60–64.

(7) Avery, M. A.; Mizuno, C. S.; Chittiboyina, A. G.; Kurtz, T. W.; Pershadsingh, H. A. Type 2 diabetes and oral antihyperglycemic drugs. *Curr. Med. Chem.* **2008**, *15*, 61–74.

(8) Barnett, A. H. Thiazolidinediones and cardiovascular outcomes. Br. J. Diabetes Vasc. Dis. **2008**, *8*, 45–49.

(9) Rendell, M. The role of sulphonylureas in the management of type 2 diabetes mellitus. *Drugs* **2004**, *64*, 1339–1358.

(10) Ohishi, T.; Yoshida, S. The therapeutic potential of GPR119 agonists for type 2 diabetes. *Expert Opin. Invest. Drugs* **2012**, *21*, 321–328.

(11) Toda, N.; Hao, X.; Ogawa, Y.; Oda, K.; Yu, M.; Fu, Z.; Chen, Y.; Kim, Y.; Lizarzaburu, M.; Lively, S.; Lawlis, S.; Murakoshi, M.; Nara, F.; Watanabe, N.; Reagan, J. D.; Tian, H.; Fu, A.; Motani, A.; Liu, Q.; Lin, Y.-J.; Zhuang, R.; Xiong, Y.; Fan, P.; Medina, J.; Li, L.; Izumi, M.; Okuyama, R.; Shibuya, S. Potent and orally bioavailable GPR142 agonists as novel insulin secretagogues for the treatment of type 2 diabetes. ACS Med. Chem. Lett. **2013**, *4*, 790–794.

(12) Itoh, Y.; Kawamata, Y.; Harada, M.; Kobayashi, M.; Fujii, R.; Fukusumi, S.; Ogi, K.; Hosoya, M.; Tanaka, Y.; Uejima, H.; Tanaka, H.; Maruyama, M.; Satoh, R.; Okubo, S.; Kizawa, H.; Komatsu, H.; Matsumura, F.; Noguchi, Y.; Shinohara, T.; Hinuma, S.; Fujisawa, Y.; Fujino, M. Free fatty acids regulate insulin secretion from pancreatic β cells through GPR40. *Nature* **2003**, *422*, 173–176.

(13) Briscoe, C. P.; Tadayyon, M.; Andrews, J. L.; Benson, W. G.; Chambers, J. K.; Eilert, M. M.; Ellis, C.; Elshourbagy, N. A.; Goetz, A. S.; Minnick, D. T.; Murdock, P. R.; Sauls, H. R., Jr.; Shabon, U.; Spinage, L. D.; Strum, J. C.; Szekeres, P. G.; Tan, K. B.; Way, J. M.; Ignar, D. M.; Wilson, S.; Muir, A. I. The orphan G protein-coupled receptor GPR40 is activated by medium and long chain fatty acids. *J. Biol. Chem.* **2003**, *278*, 11303–11311.

(14) Kotarsky, K.; Nilsson, N. E.; Flodgren, E.; Owman, C.; Olde, B. A human cell surface receptor activated by free fatty acids and thiazolidinedione drugs. *Biochem. Biophys. Res. Commun.* **2003**, *301*, 406–410.

(15) Shapiro, H.; Shachar, S.; Sekler, I.; Hershfinkel, M.; Walker, M. D. Role of GPR40 in fatty acid action on the β cell line INS-1E. Biochem. Biophys. Res. Commun. 2005, 335, 97–104.

(16) Fujiwara, K.; Maekawa, F.; Yada, T. Oleic acid interacts with GPR40 to induce Ca²⁺ signaling in rat islet β -cells: mediation by PLC and L-type Ca²⁺ channel and link to insulin release. *J. Physiol. Endoclinol. Metab.* **2005**, 289, E670–E677.

(17) Tan, C. P.; Feng, Y.; Zhou, Y. P.; Eiermann, G. J.; Petrov, A.; Zhou, C.; Lin, S.; Salituro, G.; Meinke, P.; Mosley, R.; Akiyama, T. E.; Einstein, M.; Kumar, S.; Berger, J. P.; Mills, S. G.; Thornberry, N. A.; Yang, L.; Howard, A. D. Selective small-molecule agonists of G protein–coupled receptor 40 promote glucose-dependent insulin secretion and reduce blood glucose in mice. *Diabetes* **2008**, *57*, 2211–2219.

(18) Burant, C. F.; Viswanathan, P.; Marcinak, J.; Cao, C.; Vakilynejad, M.; Xie, B.; Leifke, E. TAK-875 versus placebo or glimepiride in type 2 diabetes mellitus: a phase 2, randomised, doubleblind, placebo-controlled trial. *Lancet* **2012**, *379*, 1403–1411.

(19) Tsujihata, Y.; Itoh, R.; Suzuki, M.; Harada, A.; Negoro, N.; Yasuma, T.; Momose, Y.; Takeuchi, K. TAK-875, an orally available G protein-coupled receptor 40/free fatty acid receptor 1 agonist, enhances glucose-dependent insulin secretion and improves both postprandial and fasting hyperglycemia in type 2 diabetic rats. *J. Pharmacol. Exp. Ther.* **2011**, 339, 228–237.

(20) Bharate, S. B.; Nemmani, K. VS.; Vishwakarma, R. A. Progress in the discovery and development of small-molecule modulators of G-protein-coupled receptor 40 (GPR40/FFA1/FFAR1): an emerging target for type 2 diabetes. *Expert Opin. Ther. Pat.* 2009, *19*, 237–264.

(21) Holliday, N. D.; Watson, S. J.; Brown, A. J. H. Drug discovery opportunities and challenges at G protein coupled receptors for long chain free fatty acids. *Front. Endocrinol.* **2011**, *2*, 112.

(22) Negoro, N.; Sasaki, S.; Mikami, S.; Ito, M.; Suzuki, M.; Tsujihata, Y.; Ito, R.; Harada, A.; Takeuchi, K.; Suzuki, N.; Miyazaki, J.; Santou, T.; Odani, T.; Kanzaki, N.; Funami, M.; Tanaka, T.; Kogame, A.; Matsunaga, S.; Yasuma, T.; Momose, Y. Discovery of TAK-875: A potent, selective, and orally bioavailable GPR40 agonist. ACS Med. Chem. Lett. 2010, 1, 290–294.

(23) Christiansen, E.; Due-Hansen, M. E.; Urban, C.; Merten, N.; Pfleiderer, M.; Karlsen, K. K.; Rasmussen, S. S.; Steensgaard, M.; Hamacher, A.; Schmidt, J.; Drewke, C.; Petersen, R. K.; Kristiansen, K.; Ullrich, S.; Kostenis, E.; Kassack, M. U.; Ulven, T. Structure– activity study of dihydrocinnamic acids and discovery of the potent FFA1 (GPR40) agonist TUG-469. ACS Med. Chem. Lett. **2010**, *1*, 345–349.

(24) Houze, J. B.; Zhu, L.; Sun, Y.; Akerman, M.; Qiu, W.; Zhang, A. J.; Sharma, R.; Schmitt, M.; Wang, Y.; Liu, J.; Liu, J.; Medina, J. C.; Reagan, J. D.; Luo, J.; Tonn, G.; Zhang, J.; Lu, J. Y. L.; Chen, M.; Lopez, E.; Nguyen, K.; Yang, L.; Tang, L.; Tian, H.; Shuttleworth, S. J.; Lin, D. C. -H. AMG 837: A potent, orally bioavailable GPR40 agonist. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 1267–1270.

(25) Zhou, C.; Tang, C.; Chang, E.; Ge, M.; Lin, S.; Cline, E.; Tan, C. P.; Feng, Y.; Zhou, Y. P.; Eiermann, G. J.; Petrov, A.; Salituro, G.; Meinke, P.; Mosley, R.; Akiyama, T. E.; Einstein, M.; Kumar, S.; Berger, J.; Howard, A. D.; Thornberry, N.; Mills, S. G.; Yang, L. Discovery of 5-aryloxy-2,4-thiazolidinediones as potent GPR40 agonists. *Bioorg. Med. Chem. Lett.* **2010**, *20*, 1298–1301.

(26) McKeown, S. C.; Corbett, D. F.; Goetz, A. S.; Littleton, T. R.; Bigham, E.; Briscoe, C. P.; Peat, A. J.; Watson, S. P.; Hickey, D. M. B. Solid phase synthesis and SAR of small molecule agonists for the GPR40 receptor. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 1584–1589.

(27) Christiansen, E.; Hansen, S. V. F.; Urban, C.; Hudson, B. D.; Wargent, E. T.; Grundmann, M.; Jenkins, L.; Zaibi, M.; Stocker, C. J.; Ullrich, S.; Kostenis, E.; Kassack, M. U.; Milligan, G.; Cawthorne, M. A.; Ulven, T. Discovery of TUG-770: A highly potent free fatty acid receptor 1 (FFA1/GPR40) agonist for treatment of type 2 diabetes. *ACS Med. Chem. Lett.* **2013**, *4*, 441–445.

(28) Brown, S. P.; Dransfield, P. J.; Vimolratana, M.; Jiao, X.; Zhu, L.; Pattaropong, V.; Sun, Y.; Liu, J.; Luo, J.; Zhang, J.; Wong, S.; Zhung, R.; Guo, Q.; Li, F.; Medina, J. C.; Swaminath, G.; Lin, D. C. -H.; Houze, J. B. Discovery of AM-1638: a potent and orally bioavailable GPR40/FFA1 full agonist. ACS Med. Chem. Lett. **2012**, 3, 726–730.

(29) Takano, R.; Yoshida, M.; Inoue, M.; Honda, T.; Nakashima, R.; Matsumoto, K.; Yano, T.; Ogata, T.; Watanabe, N.; Toda, N. Discovery of 3-aryl-3-ethoxypropanoic acids as orally active GPR40 agonists. *Bioorg. Med. Chem. Lett.* **2014**, *24*, 2949–2953. (30) The screening package "Lead Profiling Screen" provided by MDS Pharma Services. The receptors are shown in Supporting Information Table S5.

(31) Fujii, A.; Hashiguchi, S.; Uematsu, N.; Ikariya, T.; Noyori, R. Ruthenium(II)-catalyzed asymmetric transfer hydrogenation of ketones using a formic acid-triethylamine mixture. *J. Am. Chem. Soc.* **1996**, *118*, 2521–2522.

(32) Brooks, D. W.; Lu, L. D.-L.; Masamune, S. C-Acylation under virtually neutral conditions. *Angew. Chem., Int. Ed.* **1979**, *18*, 72–74.

(33) Nakashima, R.; Yano, T.; Ogawa, J.; Tanaka, N.; Toda, N.; Yoshida, M.; Takano, R.; Inoue, M.; Honda, T.; Kume, S.; Matsumoto, K. Potentiation of insulin secretion and improvement of glucose intolerance by combining a novel G protein-coupled receptor 40 agonist DS-1558 with glucagon-like peptide-1 receptor agonists. *Eur. J. Pharmacol.* **2014**, *737*, 194–201.